WATER COLUMN TIME SERIES PROTOCOL
FLOW CYTOMETRY (CRYOVIAL) SAMPLES PREPARATION
Moorea Coral Reef LTER

Get Together

9% paraformaldehyde, stored in freezer, 1 tube removed and thawed in fridge day prior
2 ml cryovials
Falcon tube sampled at each depth and station.

Procedure:

1. Place 1.8 ml of seawater sample from the falcon tube (shake first) into each of 2 labeled 2 ml cryovials: 2 replicates for each station and depth (use 1000 µl pipette at 900 µl twice-you can use same pipette tip) (blue cryovial box)
2. Fix with 50 µl of 9% paraformaldehyde. (use same pipette as C14)
3. Once finished, shake entire cryovial box
4. Freeze at -80° C (clear cryovial box)
5. Rinse falcon tubes with DI water-will use these tubes next time
6. Return to UCSB on dry ice or -80°C ice and take to the Carlson Lab for later analysis using flow cytometry. It is very important that these samples stay as close to -80 as possible when bringing home. Pack cryovials in the middle of cooler, surrounded by TOC bottles, etc.

Written by A.Alldrege. Updated March 2011.

(May 2011 'Preparation' appended to title to distinguish this sample preparation protocol from the analysis protocol.)